

# Genomic DNA from tissue

**User manual** 

NucleoSpin® 96 Tissue Core Kit

February 2012/Rev.06



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# 1 Components

# 1.1 Kit contents

	Nu	cleoSpin <sup>®</sup> 96 Tis	sue
	2 x 96 preps	4 x 96 preps	24 x 96 preps
REF	740741.2	740741.4	740741.24 <sup>1</sup>
Lysis Buffer T1	50 mL	100 mL	6 x 100 mL
Binding Buffer BQ1	50 mL	100 mL	6 x 100 mL
Wash Buffer B5 (Concentrate) <sup>2</sup>	100 mL	2 x 100 mL	12 x 100 mL
Wash Buffer BW	125 mL	2 x 125 mL	12 x 125 mL
Elution Buffer BE <sup>3</sup>	50 mL	100 mL	6 x 100 mL
Proteinase K (lyophilized) <sup>2</sup>	2 x 75 mg	4 x 75 mg	24 x 75 mg
Proteinase Buffer PB	8 mL	15 mL	6 x 15 mL
NucleoSpin® Tissue Binding Plates (green rings)	2	4	24
Round-well Blocks <sup>4</sup>	2	4	24
MN Square-well Blocks	2	4	24
MN Wash Plates <sup>5</sup>	2	4	24
Rack of Tube Strips <sup>6</sup>	2	4	24
Cap Strips	24	48	288
Self-adhering PE Foil	5	10	60
User manual	1	1	6

 $<sup>^{1}</sup>$  The kit for 24 x 96 preparations (REF 740741.24) consists of 6 x REF 740741.4.

<sup>&</sup>lt;sup>2</sup> For preparation of working solutions and storage conditions, see section 3.

<sup>&</sup>lt;sup>3</sup> Elution Buffer BE: 5 mM Tris/HCl, pH 8.5

<sup>&</sup>lt;sup>4</sup> Including 12 Cap Strips for each block

 $<sup>^{\</sup>rm 5}$  For use with vacuum only

<sup>&</sup>lt;sup>6</sup> Sets of 1 rack, 12 strips with 8 tubes each, including Cap Strips

#### 1.1 Kit contents continued

	NucleoSpin <sup>®</sup> 96 Tissue Core Kit
	4 x 96 preps
REF	740454.4
Lysis Buffer T1	100 mL
Binding Buffer BQ1	100 mL
Wash Buffer B5 (Concentrate) <sup>1</sup>	2 x 100 mL
Wash Buffer BW	2 x 125 mL
Elution Buffer BE <sup>2</sup>	100 mL
Proteinase K (lyophilized) <sup>1</sup>	4 x 75 mg
Proteinase Buffer PB	15 mL
NucleoSpin® Tissue Binding Plates (green rings)	4
User manual	1

# 1.2 Reagents to be supplied by user

96–100% ethanol (for preparation of working solutions; see section 3)

For more detailed information regarding special hardware required for centrifuge or vacuum processing, please see section 2.3.

For recommended accessories for use of the flexible **NucleoSpin® 96 Tissue Core Kit** (reduced kit composition; REF 740454.4), please see section 2.4.

<sup>&</sup>lt;sup>1</sup> For preparation of working solutions and storage conditions see section 3.

<sup>&</sup>lt;sup>2</sup> Elution Buffer BE: 5 mM Tris/HCl, pH 8.5

# 2 Product description

# 2.1 The basic principle

The NucleoSpin® 96 Tissue kit is designed for the efficient isolation of high molecular weight genomic DNA from tissue samples or cells. With the NucleoSpin® 96 Tissue procedure, sample lysis is achieved by incubation of the samples in a solution containing SDS and Proteinase K. Appropriate conditions for binding of DNA to the silica membrane in the NucleoSpin® Tissue Binding Plate are created by addition of large amounts of chaotropic salt and ethanol to the lysate. The binding process is reversible and specific to nucleic acids. While DNA is kept on the silica membrane, contaminations are removed by washing with two different wash buffers. Pure genomic DNA is finally eluted under low ionic strength conditions in a slightly alkaline elution buffer.

# 2.2 Kit specifications

- NucleoSpin® 96 Tissue is designed for the rapid preparation of highly pure genomic DNA from tissue, for example, mouse and rat tails, organ tissue, or animal or bacterial cells. The purified DNA can be used directly as template for PCR, blotting, or any kind of enzymatic reactions.
- This kit provides reagents and consumables for purification of up to 40  $\mu$ g (average 20  $\mu$ g) of pure genomic DNA from up to 20 mg tissue samples with an  $A_{260}/A_{280}$  ratio between 1.8 and 1.9 and a typical concentration of 100–200 ng/ $\mu$ L.
- From up to two 0.5 cm long mouse tail tip section (age of mice: 4–6 weeks), up to 35 μg of pure genomic DNA can be prepared (typical yields: 15–25 μg).
- NucleoSpin® 96 Tissue can be processed by vacuum or in a centrifuge. The kit allow easy automation on common liquid handling instruments.
- The NucleoSpin® 96 Tissue kits allow for the purification of multiples of 96 samples. The kits are supplied with accessory plates for highest convenience. The kits are designed for manual or automated use in a centrifuge or for use with a vacuum manifold. The NucleoSpin® 96 Tissue Core Kit provides the buffers, Proteinase K and NucleoSpin® Tissue Binding Plate only. Accessory components (e.g., lysis plates, elution plates) are not provided with the core kit but can be individually selected from a variety of suitable accessories (see section 2.4 for further information). This allows highest flexibility for the user.

Kit specifications at a glance			
Parameter	NucleoSpin® 96 Tissue		
Format	96-well plates		
Processing	Manual and automated, vacuum or centrifugation		
Sample material	Up to 20 mg tissue, up to $10^6$ cultured cells, bacteria		
Typical yield	15–25 μg		
A <sub>260</sub> /A <sub>280</sub>	1.8–1.9		
Elution volume	100–200 μL		
Preparation time	60 min/plate (excl. lysis)		
Binding capacity	40 μg		

# 2.3 Required hardware

**NucleoSpin® 96 Tissue** can be processed under vacuum or with centrifugation. Certain hardware for processing is required.

#### Centrifugation

For centrifugation, a microtiterplate centrifuge is required. This centrifuge must be able to accommodate the NucleoSpin® Tissue Binding Plate stacked on a Round- or Squarewell Block and reach accelerations of  $5,600-6,000 \times g$  is required (bucket height: 85 mm).

Regarding waste collection, suitable consumables (e.g., MN Square-well Blocks) are necessary and they are not included in the kit. For the most convenient handling, without the need of emptying and reusing the MN Square-well Blocks, we recommend using six MN Square-well Blocks if two 96-well plates are processed at once (see ordering information). Alternatively, it is possible to empty the MN Square-well Blocks after every centrifugation step, reducing the amount of MN Square-well Blocks needed.

#### Vacuum processing

The NucleoSpin® 96 Tissue kit can be used with the NucleoVac 96 Vacuum Manifold (see ordering information). When using NucleoSpin® 96 Tissue with less than 96 samples, Self-adhering PE Foil (see ordering information) should be used in order to close and protect non-used wells of the NucleoSpin® Tissue Binding Plate and thus quarantee proper vacuum.

Establish a reliable vacuum source for the NucleoVac 96 Vacuum Manifold. The manifold may be used with a vacuum pump, house vacuum, or water aspirator. We recommend a vacuum of -0.2 to -0.4 bar (reduction of atmospheric pressure). The use of the NucleoVac Vacuum Regulator (see ordering information) is recommended.

Alternatively, adjust the vacuum so that during the purification the sample flows through the column with a rate of 1–2 drops per second. Depending on the amount of sample being used, the vacuum times may need to be increased for complete filtration.

Additionally, a suitable centrifuge for sample preparation steps may be required.

For general consumables and equipment needed, please see section 1.2.

# 2.4 Accessories supplied for use of the NucleoSpin® 96 Tissue Core Kit

The **NucleoSpin® 96 Tissue Core Kit** provides buffers, Proteinase K, and NucleoSpin® Tissue Binding Plates. Accessory plates (e.g., lysis plates, elution plates) are not provided with the core kit. The reduced kit composition along with a variety of separately available accessories, allow optimal adjustment of the kit to individual user needs. The user can select additional consumables according to his/her requirements for highest flexibility.

For use of **NucleoSpin® 96 Tissue Core Kit**, follow the standard protocols (see section 5.1 and 5.2).

Recommended accessories for use of the **NucleoSpin® 96 Tissue Core Kit** are available from MACHEREY-NAGEL (see ordering information).

Protocol step	Suitable consumables, not supplied with the core kits	Remarks
Lyse samples	4 x Rack of -well Block with Cap Strips per 4 x 96 preps	Foe sample lysis
	or  4 x Rack of Tube Strips with Cap Strips per 4 x 96 preps	

Protocol step	Suitable consun supplied with th	Remarks
Adjust binding conditions	48 x Cap Strips per 4 x 96 preps	When using Round-well Block or Tube Strips for lysis, new Cap Strips are required for sealing of wells after adding Buffer BQ1 and ethanol.
	4 x MN Square- well Block per 4 x 96 preps	Recommended for automated processing only
Bind DNA to the membrane	4 x MN Wash Plate per 4 x 96 preps	MN Wash Plate minimizes the risk of cross contamination (vacuum processing only).
	2 x MN Square- well Block	For waste collection during centrifugation (reusable)
Elute DNA	4 x Rack of Tubes Strips with Cap Strips per 4 x 96 preps	
	or 4 x Round-well	
	Block with Cap Strips per 4 x 96 preps	

# 2.5 Automated processing on robotic platforms

**NucleoSpin® 96 Tissue** can be used fully automated on many common laboratory workstations. For the availability of scripts and general considerations about adapting **NucleoSpin® 96 Tissue** on a certain workstation, please contact MN. Full processing under vacuum enables complete automation without the need for centrifugation steps for drying of the binding membrane or for elution. However, a final elution step by centrifugation is recommended in order to achieve higher concentrated eluted DNA.

The risk of cross-contamination is reduced by optimized vacuum settings during the elution step and by the improved shape of the outlets of the NucleoSpin® Tissue Binding Plate.

Drying of the NucleoSpin® Tissue Binding Plates under vacuum is sufficient because the bottom of the plate is protected by the MN Wash Plate during the washing steps. As a result, it is recommended to integrate the MN Wash Plate into the automated procedure to protect against these wash buffer residues. The MN Frame (see ordering information) can be used to position the disposable MN Wash Plate inside the vacuum chamber. This also reduces the risk of cross-contamination, as common metal adaptors tend to get contaminated by gDNA. Thorough cleaning of the vacuum chamber is recommended after each run to prevent DNA-containing aerosols from forming.

Visit MN online at *www.mn-net.com* or contact your local MACHEREY-NAGEL distributor for technical support regarding hardware, software, setup instructions, and selection of the protocol. Several application notes of the **NucleoSpin® 96 Tissue** kit on various liquid handling instruments can also be found at *www.mn-net.com* at Bioanalysis/Literature.

# 2.6 Elution procedures

It is possible to adjust the elution method and the volume of the elution buffer to the subsequent application of interest. In addition, to the standard method described in the protocols (recovery rate about 70–90 %) there are several modifications possible. Use elution buffer preheated at 70  $^{\circ}$ C for one of the following procedures:

- High yield: Perform two elution steps with the volume indicated in the individual protocol. About 90–100 % of bound nucleic acids can be eluted.
- High concentration: Perform one elution step with only 60% of the volume indicated in the individual protocol. Concentration of DNA will be about 30% higher than with the standard elution procedure. Maximum yield of bound nucleic acids is about 80%.
- High yield and high concentration: Apply half the volume of elution buffer as indicated in the individual protocol, incubate for 3 min and centrifuge. Apply a second aliquot of elution buffer, incubate and centrifuge again. Thus, about 85–100% of bound nucleic acids are eluted in the standard elution volume at a high concentration.

 Convenient elution: For convenience, elution buffer of ambient temperature may be used. This will result in a slightly lower yield (approximately 20%) compared to elution with heated elution buffer.

Elution may also be performed with Tris-EDTA-buffer (TE) of pH equal or higher than 8. This will increase DNA stability during long term or multi-use storage at  $4\,^{\circ}$ C (or ambient temperature) by inhibiting omnipresent DNases. However, EDTA interferes, depending on the final concentration, with certain downstream applications.

For optimal performance of isolated DNA in downstream applications, we recommend eluting with the supplied elution buffer and storing it, especially long term, at - 20 °C. Several freeze-thaw cycles will not interfere with most downstream applications.

Performance of long-range PCR (e.g., > 10 kb) or the detection limit of trace amount of DNA species, may be reduced after multiple freeze-thaw cycles or prolonged storage of eluted DNA at 4 °C or room temperature. This is due to shearing of DNA or adsorption to surfaces.

Due to the dead volume of the silica membrane, please note that the difference between the dispensed elution buffer volume and the recovered elution buffer volume containing genomic DNA is approximately 20  $\mu$ L (recovered elution volume = dispensed elution volume - 20  $\mu$ L).

# 3 Storage conditions and preparation of working solutions

Attention: Buffer BQ1 and BW contain chaotropic salts. Wear gloves and goggles!

CAUTION: Buffers BQ1 and BW contain guanidine hydrochloride which can form highly reactive compounds when combined with bleach (sodium hypochlorite). DO NOT add bleach or acidic solutions directly to the sample-preparation waste.

#### Storage conditions:

All components of the NucleoSpin® 96 Tissue kits should be stored at room temperature (18–25 °C) for a maximum of 1 year. Storage at lower temperatures may cause precipitation of salts. If a salt precipitation is observed, incubate the bottle at 30–40 °C for some minutes and mix well until all of the precipitate is redissolved. The performance of the kits is not affected by the salt precipitates.

Before starting any NucleoSpin® 96 Tissue protocol, prepare the following:

- Wash Buffer B5: Add the indicated volume of ethanol (96–100 %) to Buffer B5 Concentrate before use. Mark the label of the bottle to indicate that ethanol was added. Store Wash Buffer B5 at room temperature (18–25 °C) for up to one year.
- Before first use of the kit, add the indicated volume of Proteinase Buffer PB to lyophilized Proteinase K. Proteinase K solution is stable at - 20 °C for up to 6 months.

	N	lucleoSpin <sup>®</sup> 96 Tissu	е
	2 x 96 preps	4 x 96 preps	24 x 96 preps
REF	740741.2	740741.4	740741.24
Wash Buffer B5 (Concentrate)	100 mL Add 400 mL ethanol	2 x 100 mL Add 400 mL ethanol to each bottle	12 x 100 mL Add 400 mL ethanol to each bottle
Proteinase K (lyophilized)	2 x 75 mg Add 2.6 mL Proteinase Buffer PB to each vial	4 x 75 mg Add 2.6 mL Proteinase Buffer PB to each vial	24 x 75 mg Add 2.6 mL Proteinase Buffer PB to each vial

	NucleoSpin <sup>®</sup> 96 Tissue Core Kit
	4 x 96 preps
REF	740454.4
Wash Buffer B5 (Concentrate)	2 x 100 mL Add 400 mL ethanol to each bottle
Proteinase K (lyophilized)	4 x 75 mg Add 2.6 mL Proteinase Buffer to each vial

# 4 Safety instructions – risk and safety phrases

The following components of the NucleoSpin® 96 Tissue and NucleoSpin® 96 Tissue Core kits contain hazardous contents.

Wear gloves and goggles and follow the safety instructions given in this section.

# 4.1 Risk and safety phrases

Component	Hazard contents	Hazard symbol	Risk phrases	Safety phrases
Inhalt	Gefahrstoff	Gefahrstoff- symbol	R-Sätze	S-Sätze
BQ1	Guanidine hydrochloride 50–66 % Guanidinhydrochlorid 50–66 %	<b>X</b> Xn*	R 22-36/38	S 26- 37/39
BW	Guanidine hydrochloride 36–50 % + isopropanol 20–50 % Guanidinhydrochlorid 36–50 % + Isopropanol 20–50 %	<b>X</b> Xn*	R 10-22- 36	S 16-26- 39
Proteinase K	Proteinase K, lyophilized Proteinase K, lyophilisiert	<b>X</b> Xn	R 36/37/38- 42	S 22-24- 26-36/37

#### Risk phrases

R 10	Flammable. Leichtentzündlich
R 22	Harmful if swallowed. Gesundheitsschädlich beim Verschlucken.
R 36	Irritating to the skin. Reizt die Haut.
R 36/38	Irritating to eyes and skin. Reizt die Augen und die Haut.
R 36/37/38	Irritating to eyes, respiratory system, and skin. Reizt die Augen, die Atmungsorgane und die Haut.
R 42	May cause sensitization by inhalation. Sensibilisierung durch Einatmen möglich.

#### Safety phrases

S 16	Keep away from sources if ignition – No smoking! Von Zündquellen fernhalten – Nicht rauchen!
S 22	Do not breathe dust. Staub nicht einatmen.
S 24	Avoid contact with the skin.  Berührung mit der Haut vermeiden.

<sup>\*</sup> Hazard labeling not necessary if quantity per bottle below 125 g or mL (certificate of exemption according to 67/548/EEC Art. 25, 1999/45/EC Art. 12 and German GefStoffV § 20 (3) and TRGS 200 7.1). For further information see Material Safety Data Sheet.

# Safety phrases

S 26	In case of contact with eyes, rinse immediately with plenty of water and seek medical advice.  Bei Berührung mit den Augen sofort gründlich mit Wasser abspülen und Arzt konsultieren.
S 36/37	Wear suitable protective clothing and gloves. Bei der Arbeit geeignete Schutzhandschuhe und Schutzkleidung tragen.
S 37/39	Wear suitable gloves and eye / face protection. Bei der Arbeit geeignete Schutzhandschuhe und Schutzbrille / Gesichtsschutz tragen.
S 39	Wear eye / face protection. Schutzbrille / Gesichtsschutz tragen.

## 4.2 GHS classification

Only harmful features need not be labeled with H and P phrases until 125 mL or 125 g. Mindergefährliche Eigenschaften müssen bis 125 mL oder 125 g nicht mit H- und P-Sätzen gekennzeichnet werden.

Component	Hazard contents	GHS symb	ool	Hazard phrases	Precaution phrases
Inhalt	Gefahrstoff	GHS Symbo	ol	H-Sätze	P-Sätze
BQ1	Guanidine hydrochloride 50–66 % Guanidinhydrochlorid 50–66 %	(I)	Warning  Achtung	302, 315, 319	280, 301+312, 302+352, 305+351+338, 330, 332+313, 337+313
BW	Guanidine hydrochloride 36–50 % + isopropanol 20–50 % Guanidinhydrochlorid 36–50 % + Isopropanol 20–50 %	<b>\$</b>	Warning  Achtung	226, 302, 319	210, 233, 280, 301+312, 305+351+338, 330, 337+313, 403+235
Proteinase K	Proteinase K, lyophilized Proteinase K, lyophilisiert	<b>♦</b> ◆	Danger Gefahr	315, 319, 334, 335	261, 280, 302+352, 304+340, 305+351+338, 312, 332+313, 337+313, 342+311, 403+233

#### **Hazard phrases**

H 226	Flammable liquid and vapour. Flüssigkeit und Dampf entzündbar.
H 302	Harmful if swallowed. Gesundheitsschädlich bei Verschlucken.
H 315	Causes skin irritation. Verursacht Hautreizungen.
H 319	Causes serious eye irritation. Verursacht schwere Augenreizung.

#### **Hazard phrases**

H 334 May cause allergy or asthma symptoms or breathing difficulties if inhaled.

Kann bei Einatmen Allergie, asthmaartige Symptome oder Atembeschwerden verursa-

Keep away from heat/sparks/open flames/hot surfaces – No smoking. Von Hitze/Funken/offener Flamme/heißen Oberflächen fernhalten. Nicht rauchen.

chen.

H 335 May cause respiratory irritation.

Kann die Atemwege reizen.

Keep container tightly closed.

Get medical advice / attention.

tor/physician.

#### **Precaution phrases**

P 210

P 233

	Behälter dicht verschlossen halten.
P 261	Avoid breathing dust.  Einatmen von Staub vermeiden.
P 280	Wear protective gloves/eye protection. Schutzhandschuhe/Augenschutz tragen.
P 301+312	IF SWALLOWED: Call a POISON CENTER or doctor/physician if you feel unwell.  BEI VERSCHLUCKEN: Bei Unwohlsein GIFTINFORMATIONSZENTRUM oder Arzt
	anrufen.
P 302+352	IF ON SKIN: Wash with plenty of soap and water. BEI KONTAKT MIT DER HAUT: Mit viel Wasser und Seife waschen.
P 304+340	IF INHALED: Remove victim to fresh air and keep at rest in a position comfortable for breathing.  BEI EINATMEN: An die frische Luft bringen und in einer Position ruhigstellen, die das Atmen erleichtert.
P 305+351+338	IF IN EYES: Rinse continuously with water for several minutes. Remove contact lenses if present and easy to do – continue rinsing.  BEI KONTAKT MIT DEN AUGEN: Einige Minuten lang behutsam mit Wasser spülen.  Vorhandene Kontaktlinsen nach Möglichkeit entfernen. Weiter spülen.
P 312	Call a POISON CENTER or doctor/physician if you feel unwell. Bei Unwohlsein GIFTINGORMATIONSZENTRUM oder Arzt anrufen.
P 330	Rinse mouth.  Mund ausspülen.
P 332+313	If skin irritation occurs: Get medical advice / attention.

For further information please see Material Safety Data Sheets (www.mn-net.com). Weiterführende Informationen finden Sie in den Sicherheitsdatenblättern (www.mn-net.com).

Store in a well ventilated place. Keep cool. Kühl an einem gut belüfteten Ort aufbewahren.

Bei Hautreizung: Ärztlichen Rat einholen / ärztliche Hilfe hinzuziehen.

Bei anhaltender Augenreizung: Ärztlichen Rat einholen / ärztliche Hilfe hinzuziehen.

If experiencing respiratory symptoms: Call a POISON CENTER or doc-

Bei Symptomen der Atemwege: Giftinformationszentrum oder Arzt anrufen.

Store in a well ventilated place. Keep container tightly closed. Behälter dicht geschlossen an einem gut belüfteten Ort aufbewahren.

P 337+313

P 342+311

P 403+233

P 403+235

# 5 Protocols

# 5.1 NucleoSpin® 96 Tissue – centrifuge processing

- · For hardware requirements, refer to section 2.3.
- · For detailed information on each step, see page 21.
- For use of the NucleoSpin® 96 Tissue <u>Core Kit</u> (REF 740454.4), refer to section 2.4 regarding recommended accessories.

#### Before starting the preparation:

- · Check if Buffer B5 and Proteinase K were prepared according to section 3.
- Set incubator or oven to 56 °C.
- Preheat Flution Buffer BF to 70 °C.

# Protocol-at-a-glance

1	Prepare samples	2 x 0.5 cm mouse tail or
		up to 20 mg tissue, 10 <sup>6</sup> cultured cells, or bacteria
2	Lyse samples	180 μL T1
		25 μL Proteinase K
		Mix
		56 °C, ≥ 6 h
3	Adjust DNA binding conditions	200 μL BQ1
		200 μL ethanol (96-100 %)
		Mix
4	Transfer lysates to NucleoSpin® Tissue Binding Plate	
5	Bind DNA to silica membrane of the NucleoSpin® Tissue Binding Plate	5,600 x <i>g,</i> 10 min

6	Wash silica membrane	500 μL BW
		5,600 x <i>g,</i> 2 min
		700 µL B5
		5,600 x <i>g,</i> 4 min
7	Dry silica membrane	70°C, 10 min
8	Elute DNA	100 μL BE (70 °C)
		5,600 x <i>g</i> , 2 min
		Optional: Repeat elution step once.

# **Detailed protocol**

- For hardware requirements, refer to section 2.3.
- For use of the NucleoSpin<sup>®</sup> 96 Tissue <u>Core Kit</u> (REF 740454.4), refer to section 2.4 regarding recommended accessories.

#### Before starting the preparation:

- · Check if Buffer B5 and Proteinase K were prepared according to section 3.
- Set incubator or oven to 56 °C.
- Preheat Elution Buffer BE to 70 °C.

#### 1 Prepare samples

For each preparation, cut up to two 0.5 cm pieces (20 mg) of mouse tail into appropriate lysis tubes or plates. If preparing DNA from rat tails, one 0.5 cm piece is sufficient. Tissue samples should not exceed 20 mg, cultured cells and bacteria should not exceed 10<sup>6</sup> cells

#### 2 Lyse samples

Prepare a Proteinase K working solution: For each sample, mix  $25 \,\mu\text{L}$  **Proteinase K** with  $180 \,\mu\text{L}$  **Buffer T1** and vortex. Transfer  $200 \,\mu\text{L}$  of the resulting solution to each well of the Round-well Block containing the samples. Close the wells with Cap Strips. Mix by vigorous shaking for  $10-15 \, \text{s}$ . Spin briefly  $(15 \, \text{s}; 1,500 \, \text{x} \, g)$  to collect any sample at the bottom of the wells.

The samples must be submerged in the solution. Never prepare the Proteinase K working solution more than 15 min before addition to the samples. Proteinase K tends to self digestion when incubated in Buffer T1 without substrate.

Incubate the Round-well Block containing the samples at **56** °C for **at least 6 h** (for mammalian cells reduce incubation to 10 min, bacterial cells may require pre-lysis with, e.g., lysozyme) or overnight until the samples are completely lysed. For optimal lysis, mix occasionally during incubation. Place a weight on top of the Round-well Block in order to prevent the Cap Strips from popping off occasionally.

After lysis, set the incubator to 70 °C for the membrane drying step.

Centrifuge the Round-well Block (15 s;  $1,500 \times g$ ) to collect any condensate from the Cap Strips. Remove Cap Strips.

Residual hair and/or bones in the lysate can be removed by centrifugation (2 min;  $5,600-6,000 \times g$ ) and transfer of the supernatant to a new Round-well Block (not supplied with the kit).

#### 3 Adjust DNA binding conditions

Add 200  $\mu$ L Buffer BQ1 and 200  $\mu$ L 96–100 % ethanol to each sample. Again, take care not to moisten the rims of the individual wells while dispensing the buffer. Close the individual wells with new Cap Strips (supplied). Mix by vigorous shaking for 10–15 s. Spin briefly (10 s; 1,500 x g) to collect any sample from the Cap Strips.

Ethanol and Buffer BQ1 can be premixed before addition to the samples, if the mixture is to be used during the next 3 months. Never centrifuge at higher g-forces or for longer periods as DNA will precipitate.

Place a NucleoSpin® Tissue Binding Plate on a MN Square-well Block.

If using more than one plate, label the plates for later identification. The use of a second plate placed on a MN Square-well Block avoids the need to balance the centrifuge.

#### 4 Transfer lysates

Remove the first Cap Strip and transfer the lysates resulting from step 2 carefully from the Round-well Block into the wells of the NucleoSpin® Tissue Binding Plate. Continue with the next eight samples. Do not moisten the rims of the individual wells while dispensing the samples – moistened rims may cause cross contamination during centrifugation. After transfer, seal the openings of the plate with Self-adhering PE Foil.

For transfer of the lysate from the Round-well Block to the NucleoSpin® Tissue Binding Plate, we recommend use of an electronic eight-channel pipetting device with extra long tips capable of holding more than 650 µL.

#### 5 Bind DNA to silica membrane

Place the MN Square-well Blocks with NucleoSpin<sup>®</sup> Tissue Binding Plates onto the centrifuge carriers and insert them into the rotor buckets. Centrifuge at  $5,600-6,000 \times g$  for 10 min.

Typically, the lysates will have passed through the silica membrane within a few minutes. The centrifugation process can be extended to 20 min, if the lysates have not passed completely.

#### 6 Wash silica membrane

#### 1<sup>st</sup> wash

Remove the Self-adhering PE Foil and add 500  $\mu$ L Buffer BW to each well of the NucleoSpin<sup>®</sup> Tissue Binding Plate. Seal the plate with a new Self-adhering PE Foil and centrifuge again at 5,600–6,000 x q for 2 min.

## 2<sup>nd</sup> wash

Remove the Self-adhering PE Foil and add **700**  $\mu$ L **Buffer B5** to each well of the NucleoSpin<sup>®</sup> Tissue Binding Plate. Seal the plate with a new Self-adhering PE Foil and centrifuge again at **5,600–6,000**  $\times$  g for **4 min**.

During this step, as much ethanolic Buffer B5 as possible is removed by centrifugation.

#### 7 Dry silica membrane

Remove the Self-adhering PE Foil and place the NucleoSpin<sup>®</sup> Tissue Binding Plate on an opened Rack of Tube Strips. Place it in an incubator for **10 min** at **70 °C** to evaporate residual ethanol.

Removal of ethanol by evaporation at 70 °C is more effective than prolonged centrifugation.

Note: The ethanol in Buffer B5 may inhibit enzymatic reactions and should be removed completely before eluting DNA.

#### 8 Elute DNA

Dispense 100  $\mu$ L preheated Buffer BE (70 °C) to each well of the NucleoSpin® Tissue Binding Plate. Dispense the buffer directly onto the membrane. Incubate at room temperature for 1 min. Centrifuge at 5,600–6,000 x g for 2 min. Repeat elution step once. Remove the NucleoSpin® Tissue Binding Plate from the Rack of Tube Strips by lifting the plate at one side carefully as the Tube Strips may stick to the outlets of the NucleoSpin® Tissue Binding Plate. For alternative elution procedures see section 2.3.

If elution in small volume tubes is desired, place a 96 PCR plate (not supplied) on top of a Round-well Block or a Rack of Tube Strips and elute into the PCR plate.

# 5.2 NucleoSpin® 96 Tissue – vacuum processing

- · For hardware requirements, refer to section 2.3.
- For detailed information regarding the vacuum manifold setup, see page 26.
- · For detailed information on each step, see page 27.
- For use of the NucleoSpin® 96 Tissue <u>Core Kit</u> (REF 740454.4), refer to section 2.4 regarding recommended accessories.

#### Before starting the preparation:

- Check if Buffer B5 and Proteinase K were prepared according to section 3.
- Set incubator or oven to 56 °C.
- Preheat Elution Buffer BE to 70 °C.

# Protocol-at-a-glance

1	Prepare samples	2 x 0.5 cm mouse tail or up to 20 mg tissue, 10 <sup>6</sup> cultured cells, or bacteria
2	Lyse samples	180 μL T1
		25 μL Proteinase K
		Mix
		56 °C, ≥ 6 h
3	Adjust DNA binding conditions	200 μL BQ1
		200 μL ethanol (96–100%)
		Mix
		Prepare the NucleoVac 96 Vacuum Manifold
4	Transfer lysates to NucleoSpin® Tissue Binding Plate	
5	Bind DNA to silica membrane of the NucleoSpin® Tissue Binding Plate	-0.2 bar∗, 5 min

<sup>\*</sup> Reduction of atmospheric pressure

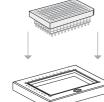
i	Wash silica membrane	600 μL BW
		900 μL B5
		900 μL B5
		-0.2 bar*, 5 min each step
		Remove MN Wash Plate
7	Dry silica membrane	- 0.6 bar∗, 10 min
3	Elute DNA	100 μL BE (70 °C)
		-0.4 bar*, 2 min
		Optional: Repeat elution step once

<sup>\*</sup> Reduction of atmospheric pressure

#### Setup of vacuum manifold:

Binding / Washing steps

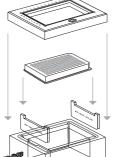




Step 4: Place the NucleoSpin® Binding Plate on top of the manifold lid.

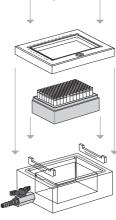


Step 4: Place the NucleoSpin® Binding Plate on top of the manifold lid.



Place the manifold lid on top of the manifold base.

Step 2: Place the MN Wash Plate in the manifold.



Step 3: Place the manifold lid on top of the manifold base.

Place the Rack of Tube

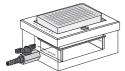
Strips in the manifold.



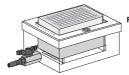


in the

Step 2:



Final setup



Final setup

# **Detailed protocol**

- For hardware requirements, refer to section 2.3.
- For detailed information regarding the vacuum manifold setup, see page 26.
- For use of the NucleoSpin<sup>®</sup> 96 Tissue <u>Core Kit</u> (REF 740454.4), refer to section 2.4 regarding recommended accessories.

#### Before starting the preparation:

- Check if Buffer B5 and Proteinase K were prepared according to section 3.
- Set incubator or oven to 56 °C.
- Preheat Elution Buffer BE to 70 °C.

#### 1 Prepare samples

For each preparation, cut up to two 0.5 cm pieces (20 mg) of mouse tail into appropriate lysis tubes or plates. If preparing DNA from rat tails, one 0.5 cm piece is sufficient. Tissue samples should not exceed 20 mg, cultured cells and bacteria should not exceed 10<sup>6</sup> cells.

## 2 Lyse samples

Prepare a Proteinase K working solution: For each sample, mix  $25 \,\mu\text{L}$  Proteinase K with  $180 \,\mu\text{L}$  Buffer T1 and vortex. Transfer 200  $\mu\text{L}$  of the resulting solution to each well of the Round-well Block containing the samples. Close the wells with Cap Strips and mix by vigorous shaking for  $10-15 \, \text{s}$ . Spin briefly  $(15 \, \text{s}; 1,500 \, \text{x} \, g)$  to collect any sample at the bottom of the wells.

The samples must be submerged in the solution. Never prepare the Proteinase K working solution more than 15 min before addition to the samples. Proteinase K tends to self digestion when incubated in Buffer T1 without substrate.

Incubate the Round-well Block containing the samples at **56** °C for **at least 6** h (for mammalian cells reduce incubation to 10 min, bacterial cells may require pre-lysis with, e.g., lysozyme) or overnight until the samples are completely lysed. For optimal lysis, mix occasionally during incubation. Place a weight on top of the Round-well Block in order to prevent the Cap Strips from popping off occasionally.

Centrifuge the Round-well Block (15 s; 1,500 x g) to collect any condensate from the Cap Strips. Remove Cap Strips.

Residual hair and/or bones in the lysate can be removed by centrifugation (2 min;  $5,600-6,000 \times g$ ) and transfer of the supernatant to a new Round-well Block (not supplied with the kit).

<sup>\*</sup> Reduction of atmospheric pressure

#### 3 Adjust DNA binding conditions

Add 200  $\mu$ L Buffer BQ1 and 200  $\mu$ L 96–100 % ethanol to each sample. Again, take care not to moisten the rims of the individual wells while dispensing the buffer. Close the individual wells with new Cap Strips (supplied). Mix by vigorous shaking for 10–15 s. Spin briefly (10 s; 1,500 x g) to collect any sample from the Cap Strips.

Ethanol and Buffer BQ1 can be premixed before addition to the samples, if the mixture is to be used up during the next 3 months. Never centrifuge at higher g-forces or for longer periods as DNA will precipitate.

#### Prepare the NucleoVac 96 Vacuum Manifold:

Place waste tray into vacuum manifold base. Insert spacers labeled "MTP/Multi-96 plate" notched side up and place the MN Wash Plate on them. Close the manifold with the manifold lid and place a NucleoSpin® Tissue Binding Plate on top of the manifold.

#### 4 Transfer lysates

Remove the first Cap Strip and transfer the lysates resulting from step 2 carefully from the Round-well Block into the wells of the NucleoSpin® Tissue Binding Plate. Continue with the next eight samples. Do not moisten the rims of the individual wells while dispensing the samples – moistened rims may cause cross-contamination.

For transfer of the lysate from the Round-well Block to the NucleoSpin® Tissue Binding Plate, we recommend use of an electronic eight-channel pipetting device with extra long tips capable of holding more than 650 µL.

#### 5 Bind DNA to silica membrane

Apply vacuum until all lysates have passed through the wells of the NucleoSpin<sup>®</sup> Tissue Binding Plate (-0.2 bar\*; 5 min). Release the vacuum.

<sup>\*</sup> Reduction of atmospheric pressure

#### 6 Wash silica membrane\*

#### 1<sup>st</sup> wash

Add **600 μL Buffer BW** to each well of the NucleoSpin® Tissue Binding Plate. Apply vacuum **(-0.2 bar\*\***; **5 min)** until all buffer has passed through the wells of the NucleoSpin® Tissue Binding Plate. Release the vacuum.

## 2<sup>nd</sup> wash

Add **900 µL Buffer B5** to each well of the NucleoSpin® TissueBinding Plate. Apply vacuum **(-0.2 bar\*\***; **5 min)** until all buffer has passed through the wells of the NucleoSpin® Tissue Binding Plate. Release the vacuum.

#### 3<sup>rd</sup> wash

Add **900 µL Buffer B5** to each well of the NucleoSpin® Tissue Binding Plate. Apply vacuum **(-0.2 bar\*\***; **5 min)** until all buffer has passed through the wells of the NucleoSpin® Tissue Binding Plate. Release the vacuum.

#### Remove MN Wash Plate

After the final washing step, close the valve, release the vacuum and remove the NucleoSpin<sup>®</sup> Tissue Binding Plate. Put it on a clean paper towel to remove residual EtOH-containing wash buffer. Remove manifold lid, MN Wash Plate, and waste container from the vacuum manifold.

#### 7 Dry silica membrane

Insert the NucleoSpin® Tissue Binding Plate into the lid, and close the manifold. Apply maximum vacuum (at least -0.6 bar\*\*) for 10 min to dry the membrane completely. This step is necessary to eliminate traces of ethanol.

<u>Note</u>: The ethanol in Buffer B5 inhibits enzymatic reactions and has to be removed completely before eluting DNA.

Finally, release the vacuum.

<sup>\*</sup> Buffer volumes are increased compared to processing under centrifugation to improve washing efficiency under vacuum.

<sup>\*\*</sup> Reduction of atmospheric pressure

#### 8 Elute DNA

Insert spacers "Microtube rack" into the NucleoVac Vacuum Manifold's short sides. Place a Rack of Tube Strips onto the spacer. Close the vacuum manifold and place the NucleoSpin® Tissue Binding Plate on top. Dispense 100 µL preheated Buffer BE onto the membrane. Incubate for 3 min at room temperature. Apply vacuum for elution (-0.4 bar\*; 2 min). Release the vacuum and repeat the elution step once. For alternative elution procedures see section 2.3.

Finally, close the Tube Strips with Cap Strips for storage.

Centrifuge Rack of Tube Strips shortly to collect all sample at the bottom of the Tube Strips.

<sup>\*</sup> Reduction of atmospheric pressure

# 6 Appendix

# 6.1 Troubleshooting

#### **Problem**

#### Possible cause and suggestions

#### Incomplete lysis

- Sample has not completely been submerged during heat incubation. Cut samples into small pieces. Mix well. Be sure that the samples are fully submerged in Buffer T1/Proteinase K mixture. Incubate until the samples are completely lysed.
- Buffer T1 and Proteinase K have been premixed more than 15 min before addition to the substrate. Proteinase K tends to self digestion under optimal reaction conditions in Buffer T1 without substrate

# No or poor DNA yield

#### Reagents not applied properly

 Prepare Buffer B5 and Proteinase K solution according to instructions (see section 3). Add Buffer BQ1 and ethanol to the lysates before loading them to the wells of the NucleoSpin<sup>®</sup> Tissue Binding Plate.

#### Suboptimal elution of DNA from the column

- Preheat Buffer BE to 70 °C before elution. Apply Buffer BE directly onto the center of the silica membrane.
- Elution efficiencies decrease dramatically if elution is done with buffers with pH < 7. Use slightly alkaline elution buffer like Buffer BE (pH 8.5).

#### RNA in sample

# RNA contamination

 If DNA free of RNA is desired, cool down to room temperature after lysis incubation and add 20 µL of an RNase A solution (20 mg/mL; see ordering information). Incubate for 15 min with moderate shaking.

Problem	Possible cause and suggestions
	Carry-over of ethanol
Poor performance of genomic	<ul> <li>After washing with Buffer B5, centrifuge ≥ 4 min at 5,600–6,000 x g in order to remove ethanolic Buffer B5 completely and evaporate residual ethanol by incubating the NucleoSpin<sup>®</sup> Tissue Binding Plate at 70 °C for 10 min.</li> </ul>
DNA in enzymatic	Increase vacuum drying time to 15 min.
reactions	Contamination of DNA with inhibitory substances
	<ul> <li>Do not elute DNA with TE buffer. EDTA may inhibit enzymatic reactions. Repurify DNA and elute in Buffer BE.</li> </ul>
	Too much starting material
	<ul> <li>Repeat the procedure, using two mouse tail sections of maximally 4–6 mm length. If processing rat tails, one 0.5 cm long tail tip section is sufficient.</li> </ul>
	Hair or bones left in the lysate after step 2
Clogged wells	<ul> <li>Centrifuge the Round-well Block for 3 min at 5,600–6,000 x g. Transfer lysates to a new Round-well Block without disturbing the debris pellet.</li> </ul>
	Incomplete passage of lysate in step 4
	• If no more than 300–500 $\mu$ L of lysate is remaining in the columns, continue with step 5. Through the addition of Buffer BW the sample is diluted and thus the sample will pass the column more easily.

# 6.2 Ordering information

Product	REF	Pack of
NucleoSpin® 96 Tissue	740741.2 740741.4 740741.24	2 x 96 preps 4 x 96 preps 24 x 96 preps
NucleoSpin® 96 Tissue Core Kit	740454.4	4 x 96 preps
NucleoSpin® 8 Tissue	740740 740740.5	12 x 8 preps 60 x 8 preps
NucleoSpin® 8 Tissue Core Kit	740453.4	48 x 8 preps
Buffer T1	740940.25	25 mL
Buffer BQ1	740923.1	11
Buffer B5 Concentrate (for 500 mL Buffer B5)	740921.100	100 mL
Buffer BW	740922.500	500 mL
Proteinase K	740506	100 mg
RNase A (lyophilized)	740505	100 mg
Rack of Tube Strips (1 set consists of 1 rack, 12 strips with 8 tubes each, and 12 Cap Strips)	740477 740477.24	4 sets 24 sets
Round-well Block (1 set consists of 1 Round-well Block and 12 Cap Strips)	740475 740475.24	4 sets 24 sets
MN Wash Plate	740479 740479.24	4 24
Cap Strips	740478 740478.24	48 288
NucleoVac 96 Vacuum Manifold	740681	1
NucleoVac Vacuum Regulator	740641	1
Self-adhering PE Foil	740676	50

Visit www.mn-net.com for more detailed product information.

# 6.3 Product use restriction/warranty

**NucleoSpin® 96 Tissue (Core Kit)** components are intended, developed, designed, and sold FOR RESEARCH PURPOSES ONLY, except, however, any other function of the product being expressly described in original MACHEREY-NAGEL product leaflets.

MACHEREY-NAGEL products are intended for GENERAL LABORATORY USE ONLY! MACHEREY-NAGEL products are suited for QUALIFIED PERSONNEL ONLY! MACHEREY-NAGEL products shall in any event only be used wearing adequate PROTECTIVE CLOTHING. For detailed information please refer to the respective Material Safety Data Sheet of the product! MACHEREY-NAGEL products shall exclusively be used in an ADEQUATE TEST ENVIRONMENT. MACHEREY-NAGEL does not assume any responsibility for damages due to improper application of our products in other fields of application. Application on the human body is STRICTLY FORBIDDEN. The respective user is liable for any and all damages resulting from such application.

DNA/RNA/PROTEIN purification products of MACHEREY-NAGEL are suitable for *IN-VITRO*-USES ONLY!

ONLY MACHEREY-NAGEL products specially labeled as IVD are also suitable for *IN-VITRO*-diagnostic use. Please pay attention to the package of the product. *IN-VITRO*-diagnostic products are expressly marked as IVD on the packaging.

IF THERE IS NO IVD SIGN, THE PRODUCT SHALL NOT BE SUITABLE FOR *IN-VITRO*-DIAGNOSTIC USE!

ALL OTHER PRODUCTS NOT LABELED AS IVD ARE NOT SUITED FOR ANY CLINICAL USE (INCLUDING, BUT NOT LIMITED TO DIAGNOSTIC, THERAPEUTIC AND/OR PROGNOSTIC USE).

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MACHEREY-NAGEL shall only be responsible for the product specifications and the performance range of MN products according to the specifications of in-house quality control, product documentation and marketing material.

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