

## Enterovirus R-gene™ - ref.: 69-005 🤨





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## EXTRACTION

#### - IN EXTRACTION ROOM -

- · Schedule a pre-treatment for stools.
- Homogenize IC1 and samples to be analysed:

CSF, throat, nasopharyngeal secretions. stool, cell culture.

- Add 10 µL IC1 to the appropriate volume of each sample to be extracted in 1 extraction tube.
- Add 10 µL IC1 to the appropriate volume of W0 in 1 extraction tube.
- · Perform extraction as below:

INSTRUMENT	КІТ	Sample + IC1 Volumes	SAMPLE TYPE	PROTOCOL	ELUTION VOLUME
	QIAamp® Viral RNA Mini Kit	140 µL of sample + 10 µL IC1	CSF		40 μL
			Respiratory samples. Stool.		60 µL
QIAcube			CSF	Purification of viral RNA	50 μL
QIACUDE			Respiratory samples. Stool.	from cell-free body fluids.	60 µL
MagNAPure Compact®	MagNA Pure Compact Nucleic Acid Isolation Kit I	200 μL of sample + 10 μL IC1	CSF. Respiratory samples.	Total_NA_Plasma _100_400	50 μL
Instrument	MagNA Pure Compact RNA Isolation Kit		CSF. Respiratory samples.	RNA_Blood_V3_2	50 μL
	NucliSens® Magnetic extraction reagents		CSF. Respiratory samples.	Generic	50 μL
NucliSENS® easyMAG™				Specific A + matrix	70 µL
_			Stool.	Specific A + 100 µL silica	50 μL
Versant™ kPCR Molecular System SP	Versant™ Sample Preparation 1.0	400 μL of sample + 10 μL IC1 (extract 250 μL)	CSF	Sample Preparation Protocol 5	65 μL (eluat 50 μL)

Store extracted samples and reagents at -18°C/-22°C

#### **ANCILLARY REAGENTS:**

- CELL Control r-gene™ ref.: 71-106
- Colour Compensation r-gene™ ref.: 71-103 for LightCycler® Instruments use only.

#### GLOSSARY:

IC1 Internal Control 1 W0 Water for extraction R9 Enterovirus premix RT Reverse Transcriptase **Crossing Threshold** 

Sample Extraction + Inhibition Control IC1sample IC1W0 Reference Extraction + Inhibition Control

## **AMPLIFICATION**

- IN AMPLIFICATION ROOM -

#### Validated on:

Stratagene<sup>®</sup> LightCycler® SmartCycler® ABI<sup>®</sup> Rotor-Gene™ /ersant™ kPCR AD **GREEN & YELLOV** FCTC / FAM & Cv3 FAM & VIC 530 & 560 FAM & HEX including LC480 including ABI StepOne™

#### AMPLIFICATION PROGRAM

• Enter the following program:

	STEPS		TIME	TEMPERATURE	CYCLES	FLUORESCENCE ACQUISITION
l	Reverse Transcription		30 min.	50°C	1	None
	Activation Hot Start Taq Polymerase		15 min.	95°C	1	None
	Amplification	Denaturation	10 sec.	95°C	45	None
		Annealing	40 sec.	60°C		530 + 560 nm
		Elongation	25 sec.	72°C		None

Note 1: for LightCycler® instruments, set SEEK TEMPERATURE to the temperature corresponding to the fluorescence acquisition.

Note 2: for LightCycler® instruments, add a cooling step: 30 sec. / 40°C / 1 cycle at the end of the PCR. Note 3: for ABI® instruments, select NONE in PASSIVE REFERENCE.

Note 4: for Rotor-Gene™, calibrate the signal by clicking on GAIN OPTIMISATION.

Note 5: for Stratagene® and Versant™ kPCR Molecular System AD instruments, select NONE in REFERENCE DYE.

#### **AMPLIFICATION PREPARATION**

#### Preparation of the premix

- Plan "n" number of amplification tubes taking into account:
  - ⇒ number of sample to be amplified;
  - ⇒ positive control (PC9):
  - ⇒ negative/ inhibition control (W0);
- Pipet 15 μL x (n + 1) Amplification Premix (R9).
- Add 0.1 µL x (n + 1) Reverse Transcriptase (RT).

#### Preparation of PCR assay

- Distribute 15 uL of the previous premix (R9 + RT) in amplification tubes.
- Add 10 µL extracted sample, positive control (PC9) and extracted W0.
- · Centrifuge for 15 sec, depending on the instruments.
- Place the amplification tubes in the instrument and run the program above according to the protocol of the instrument used.

### RESULTS

#### INTERPRETATION

#### Conditions for test validation:

- Negative Inhibition Control (530 nm): NO SIGNAL.
- IC1W0 (560 nm): CT ≤ 36 CYCLES.
- PC9 (530 nm): 18 ≤ CT ≤ 25 CYCLES.

#### Sample result interpretation

	EXTRACTION and	CT [IC1sample] ≤ C	T [IC1W0] + 3 cycles	CT [IC1sample] > CT [IC1W0] + 3 cycles		
	CONTROL	NON INI and correctly e		INHIBITED or badly extracted sample		
	SAMPLE	Calculated CT	Non calculated CT	Calculated CT	Non calculated CT	
	ENTEROVIRUS detection	Sample validated as positive	Sample validated as negative	Sample validated as positive	NOT VALID	

This outlined procedure does not replace the original protocol.







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