

# Multi-Center Evaluation of the VITEK<sup>®</sup>MS System for Mass Spectrometric Identification of Non-Enterobacteriaceae Gram Negative Bacilli

ASM 2013  
13-GM-A-1491

Ryhana Manji<sup>1</sup>, Maureen Bythrow<sup>1</sup>, John A. Branda<sup>2</sup>, Carey-Ann D. Burnham<sup>3</sup>, Mary Jane Ferraro<sup>2</sup>, Omai B. Garner<sup>4</sup>, Rebecca Jennemann<sup>5</sup>, Michael A. Lewinski<sup>4</sup>, A. Brian Mochon<sup>4</sup>, Gary W. Procop<sup>6</sup>, Sandra S. Richter<sup>6</sup>, Jenna A. Rychert<sup>2</sup>, Linda Sercia<sup>6</sup>, Lars F. Westblade<sup>3,7</sup> and Christine C. Ginocchio<sup>1,7</sup>

<sup>1</sup>Department of Pathology and Laboratory Medicine, North Shore-LIJ Health System Laboratories, Lake Success, NY; <sup>2</sup>Department of Pathology, Massachusetts General Hospital and Harvard Medical School, Boston, MA, USA;

<sup>3</sup>Department of Pathology & Immunology, Washington University School of Medicine, St. Louis, MO; <sup>4</sup>Department of Pathology and Laboratory Medicine, David Geffen School of Medicine at UCLA, Los Angeles, CA, USA;

<sup>5</sup>Barnes Jewish Hospital, St. Louis, MO; <sup>6</sup>Department of Clinical Pathology, Cleveland Clinic, Cleveland, OH; <sup>7</sup>Department of Pathology and Laboratory Medicine, Hofstra North Shore-LIJ School of Medicine, Hempstead, NY

## BACKGROUND AND OBJECTIVES

The accurate, rapid identification of the non-*Enterobacteriaceae* Gram negative bacilli (NEGNB) is critical for antibiotic selection, especially for multidrug resistant NEGNB such as *Acinetobacter* sp., *Pseudomonas* sp. and *Stenotrophomonas* sp. NEGNB also cause nosocomial infections and identification triggers prompt initiation of infection control measures. Matrix-assisted laser desorption ionization time-of-flight (MALDI-TOF) mass spectrometry (MS) can potentially provide NEGNB colony identification within minutes.

This multi-center study assessed the performance of VITEK<sup>®</sup>MS and version 2.0 of the knowledge base (bioMérieux, Marcy l'Etoile, France) for identifying NEGNB clinical isolates.

## MATERIALS AND METHODS

**I. Clinical Trial Sites:** This study was performed at five geographically diverse US sites (California, Ohio, Missouri, New York and Massachusetts).

**II. Strains Tested:** Unique NEGNB clinical isolates (n=558), comprising 18 genera and 33 species (*Achromobacter*: n=41, *Acinetobacter*: n=108, *Aeromonas*: n=35, *Alcaligenes*: n=12, *Bordetella*: n=25, *Brevundimonas*: n=7, *Burkholderia*: n=40, *Chryseobacterium*: n=8, *Elizabethkingia*: n=10, *Ochrobactrum*: n=10, *Pasteurella*: n=14, *Pseudomonas*: n=109, *Ralstonia*: n=10, *Rhizobium*: n=14, *Sphingobacterium*: n=15, *Sphingomonas*: n=9, *Stenotrophomonas*: n=53, *Vibrio*: n=38). *A. baumannii* complex included *A. baumannii* (n=19), *A. baumannii* complex (n=34), *A. calcoaceticus* (n=8), *A. nosocomialis* (n=4). Isolates were obtained from test sites or, for rare isolates, provided by bioMérieux.

**III. Preparation and Testing:** NEGNB, calibrant (*E. coli* ATCC 8739) and controls (*E. aerogenes* [ATCC 13048], *K. oxytoca* [ATCC 13182], *P. aeruginosa* [ATCC 10145], and *S. aureus* [ATCC 29213]) were grown on appropriate media for <72 hr prior to testing (see below). Repeat testing of an isolate with an acceptable identification was not permitted.

**IV. Interpretation:** NEGNB identification and confidence values were determined by comparison of spectra to VITEK<sup>®</sup>MS v2.0 knowledge base. VITEK<sup>®</sup>MS results were compared to 16s rRNA DNA sequencing.

## RESULTS: Table 1: VITEK MS Identification of NEGNB Isolates

Species	Isolates Tested n	Correct to Genus and Species n (%) <sup>a</sup>	Correct to Genus only n (%)	Total Correct Results <sup>b</sup> n (%)	Miss-ID to Genus and Species <sup>c</sup> n (%)	Miss-ID to Single Species only n (%)	Total Miss-ID <sup>d</sup> n (%)	Total no ID <sup>e</sup> n (%)
<i>Achromobacter denitrificans</i>	17	0	15 (88.2)	15 (88.2)	0	0	0	2 (11.8)
<i>Achromobacter xylosoxidans</i>	24	0	22 (91.7)	22 (91.7)	0	0	0	2 (8.3)
<i>Acinetobacter baumannii</i> complex <sup>a</sup>	65	56 (86.2)	0	56 (86.2)	0	0	0	9 (13.8)
<i>Acinetobacter haemolyticus</i>	6	6 (100)	0	6 (100)	0	0	0	0
<i>Acinetobacter junii</i>	11	6 (54.5)	3 (27.3)	9 (81.8)	0	1 (9.1)	1	2 (18.2)
<i>Acinetobacter lwoffii</i>	26	22 (84.6)	1 (3.8)	23 (88.5)	0	0	0	3 (11.5)
<i>Aeromonas hydrophila/caviae</i> <sup>b</sup>	25	16 (64.0)	8 (32.0)	24 (96.0)	0	2 (8.0)	2 (8.0)	1 (4.0)
<i>Aeromonas sobria</i>	10	4 (40.0)	6 (60.0)	10 (100)	0	1 (10.0)	1 (10.0)	0
<i>Alcaligenes faecalis</i> ssp. <i>faecalis</i>	12	11 (91.7)	0 (0)	11 (91.7)	1 (8.3)	0	1 (8.3)	0
<i>Bordetella bronchiseptica</i>	10	2 (20.0)	3 (30.0)	5 (50.0)	0	0	0	5 (50.0)
<i>Bordetella parapertussis</i>	6	6 (100.0)	0 (0)	6 (100)	0	0	0	0
<i>Bordetella pertussis</i>	9	6 (66.7)	3 (33.3)	9 (100)	0	0	0	0
<i>Brevundimonas diminuta</i>	7	7 (100)	0	7 (100)	0	0	0	0
<i>Burkholderia cepacia</i>	9	3 (33.3)	5 (55.6)	8 (88.9)	0	0	0	1 (11.1)
<i>Burkholderia multivorans</i>	25	24 (96.0)	0	24 (96.0)	0	0	0	1 (4.0)
<i>Burkholderia stabilis</i>	6	0 (0)	6 (100)	6 (100)	0	0	0	0 (0)
<i>Chryseobacterium indologenes</i>	8	7 (87.5)	0	7 (87.5)	0	0	0	1 (12.5)
<i>Elizabethkingia meningoseptica</i>	10	10 (100)	0	10 (100)	0	0	0	0
<i>Ochrobactrum anthropi</i>	10	10 (100)	0	10 (100)	0	0	0	0
<i>Pasteurella multocida</i>	14	14 (100)	0	14 (100)	0	0	0	0
<i>Pseudomonas aeruginosa</i>	57	55 (96.5)	0	55 (96.5)	0	0	0	2 (3.5)
<i>Pseudomonas fluorescens</i>	19	15 (78.9)	3 (15.8)	18 (94.7)	0	0	0	1 (5.3)
<i>Pseudomonas putida</i>	25	20 (80.0)	2 (8.0)	22 (88.0)	0	1 (4.0)	1 (4.0)	3 (12.0)
<i>Pseudomonas stutzeri</i>	8	8 (100)	0	8 (100)	0	0	0	0
<i>Ralstonia pickettii</i>	10	8 (80.0)	0	8 (80.0)	0	0	0	2 (20.0)
<i>Rhizobium radiobacter</i>	14	10 (71.4)	0	10 (71.4)	2 (14.3)	0	2 (14.3)	2 (14.3)
<i>Sphingobacterium multivorans</i>	5	4 (80.0)	0	4 (80.0)	0	0	0	1 (20.0)
<i>Sphingobacterium spiritivorum</i>	10	10 (100)	0	10 (100)	0	0	0	0
<i>Sphingomonas paucimobilis</i>	9	9 (100)	0	9 (100)	0	0	0	0
<i>Stenotrophomonas maltophilia</i>	53	51 (96.2)	0	51 (96.2)	1 (1.9)	0	1 (1.9)	1 (1.9)
<i>Vibrio cholerae</i>	11	10 (90.9)	0	10 (90.9)	0	0	0	1 (9.1)
<i>Vibrio parahaemolyticus</i>	16	14 (87.5)	1 (6.3)	15 (93.8)	0	0	0	1 (6.3)
<i>Vibrio vulnificus</i>	11	10 (90.9)	0	10 (90.9)	0	0	0	1 (9.1)

Total Isolates Tested	Correct to Genus and Species <sup>a</sup>	Correct to Genus only	Total Correct Results <sup>b</sup>	MisID to Genus and Species <sup>c</sup>	MisID to Single Species only	Total MisID <sup>d</sup>	Total No ID <sup>e</sup>
N= 558	434	78	512	4	5	9	42
%	77.8%	14.0%	91.8%	0.7%	0.9%	1.6%	7.5%

MisID: incorrect identification; ID: identification

<sup>a</sup> Correct ID to genus level with either 2 or 3 species listed including correct species (n=73) or without correct species (n=5); or single ID correct to genus but incorrect to species (n=5).

<sup>b</sup> Total IDs correct to genus level only (n=78) or genus and species (n=434).

<sup>c</sup> Single ID correct to genus level but incorrect to species level (n=5). Results are included in correct to genus level category.

<sup>d</sup> Total incorrect ID to genus and species levels (n=4) or single ID correct to genus level but incorrect to species level (n=5).

<sup>e</sup> Total with no ID due to either mixed genera (n=14) or 'no ID in knowledge base' (n=28)

## RESULTS: Table 3: Discordant Results

Number Isolates	Organism	VITEK MS Identification(s) Incorrect at Species Level <sup>a</sup>
1	<i>Acinetobacter junii</i>	<i>Acinetobacter haemolyticus</i>
2	<i>Aeromonas caviae</i>	<i>Aeromonas sobria</i>
1	<i>Aeromonas sobria</i>	<i>Aeromonas hydrophila/caviae</i>
1	<i>Pseudomonas putida</i>	<i>Pseudomonas viridiflava</i>
Number Isolates	Organism	VITEK MS Identification(s) Incorrect at Genus Level
1	<i>Alcaligenes faecalis</i> ssp. <i>faecalis</i>	<i>Staphylococcus aureus</i>
1	<i>Rhizobium radiobacter</i>	<i>Obesumbacterium proteus</i>
1	<i>Rhizobium radiobacter</i>	<i>Achromobacter denitrificans</i> and <i>Achromobacter xylosoxidans</i>
1	<i>Stenotrophomonas maltophilia</i>	<i>Ochrobactrum anthropi</i>

## SUMMARY AND CONCLUSIONS

### Summary

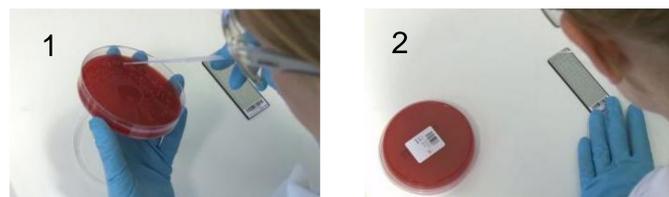
- Overall, VITEK<sup>®</sup>MS identified 91.8% of NEGNB correct to species-level (77.8%) or genus-level (14.0%).
- Overall there were 4 isolates (0.7%) misidentified to genus-level and 5 isolates (0.9%) misidentified to species-level.
- VITEK<sup>®</sup>MS called 14 NEGNB (2.5%) mixed genera and no ID was provided for the remaining 28 NEGNB (5%).

### Conclusions

- VITEK<sup>®</sup>MS provides a highly accurate and rapid identification of NEGNB.
- Time to results in minutes versus hours or days for traditional microbiologic testing allows the Laboratory to provide clinically relevant results in a time frame that can have a major impact on patient care and outcome.

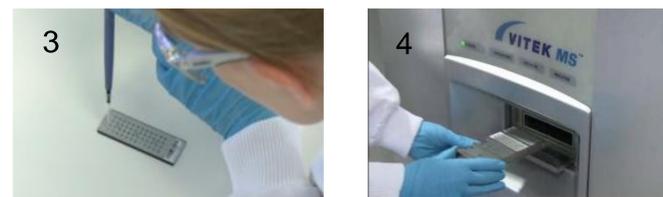
## VITEK MS SET-UP

A 1 µl loop was used to apply the bacterial isolates to wells on a disposable bar-coded target slide (Figures 1 and 2).



## VITEK MS SET-UP, DATA ANALYSIS AND INTERPRETATION

Matrix solution (α-cyano-4-hydroxycinnamic acid) was added to each well and dried (Figure 3). Slides were inserted into VITEK MS (Figure 4).



Mass spectra were generated using the VITEK<sup>®</sup>MS and mass spectra interpreted by VITEK<sup>®</sup>MS v2.0 knowledge base.



## ACKNOWLEDGEMENTS

This study was funded by bioMérieux, Durham, NC. We sincerely thank the technical staff of our laboratories. We thank David Pincus, Connie Bradford and Karen McDonald of bioMérieux, USA, for technical and statistical support.

